A practical approach to working with the BQC19 multi-*omics datasets

- Approaching a new type of data -

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SomaScan

A new tool

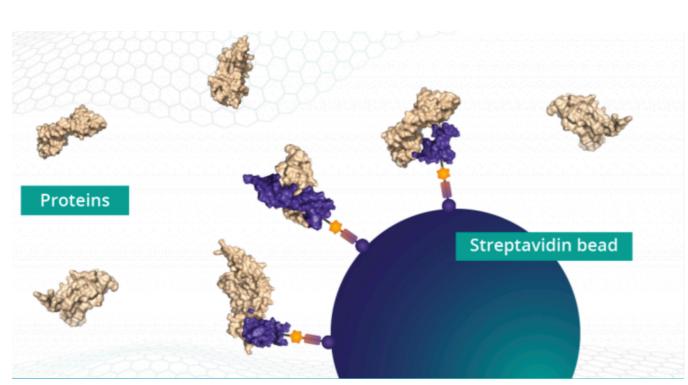
Rely on **aptamers**

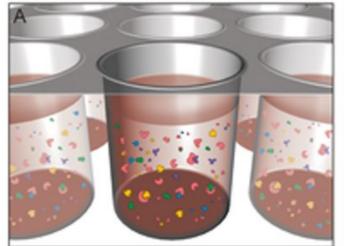
Linked to **flurophores**

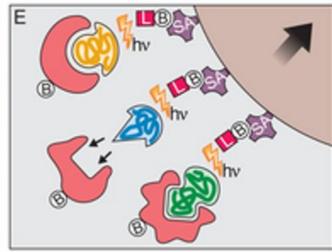
To *measure* the expression of ~5K proteins

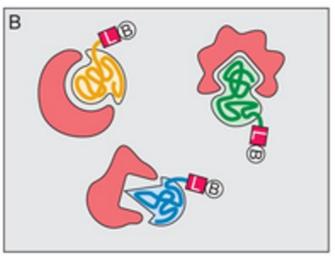
relatives measurements ⇒ normalisation & co

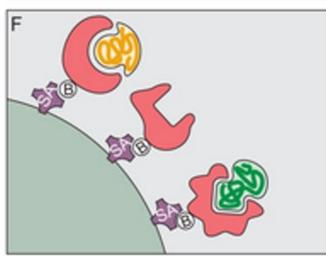
"What I wish I knew when I started using those data"

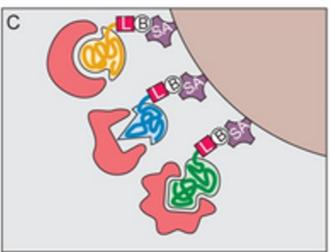




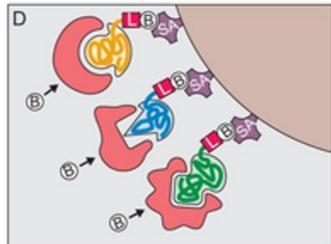


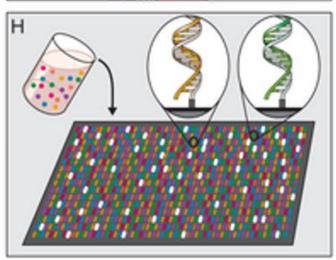




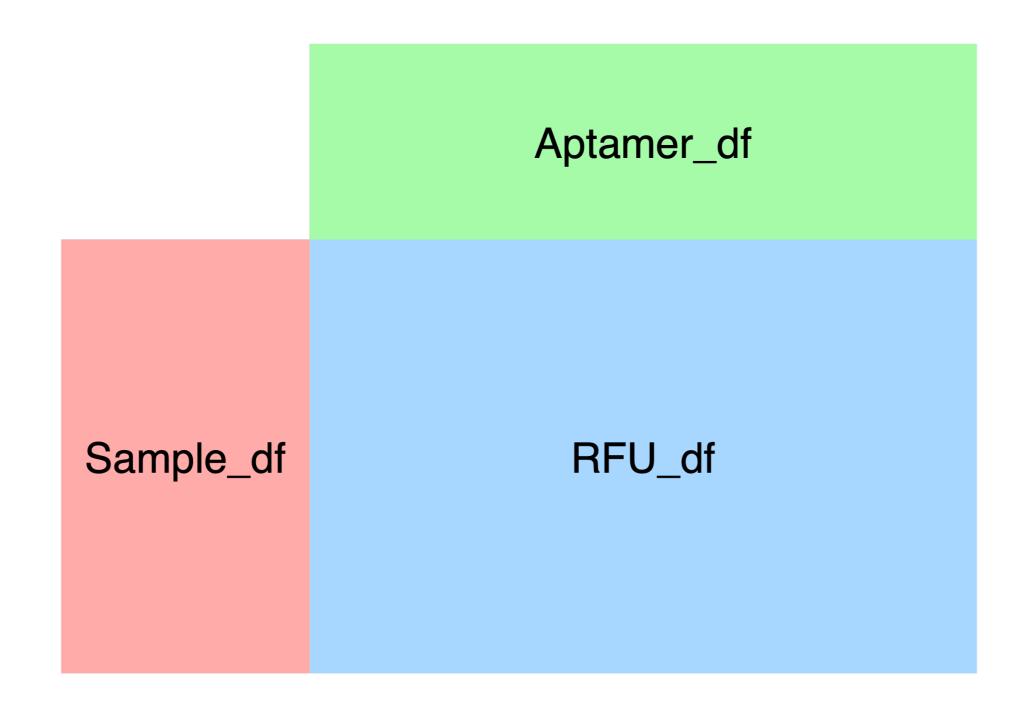




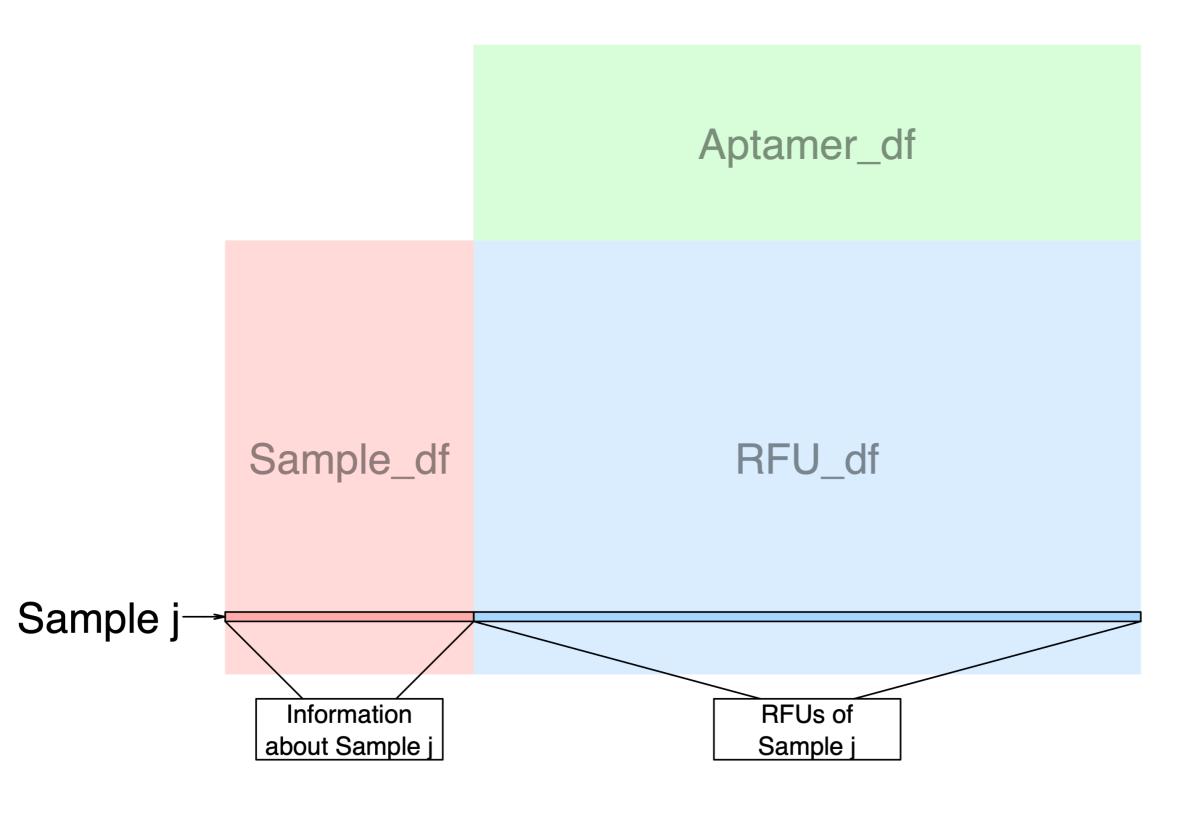




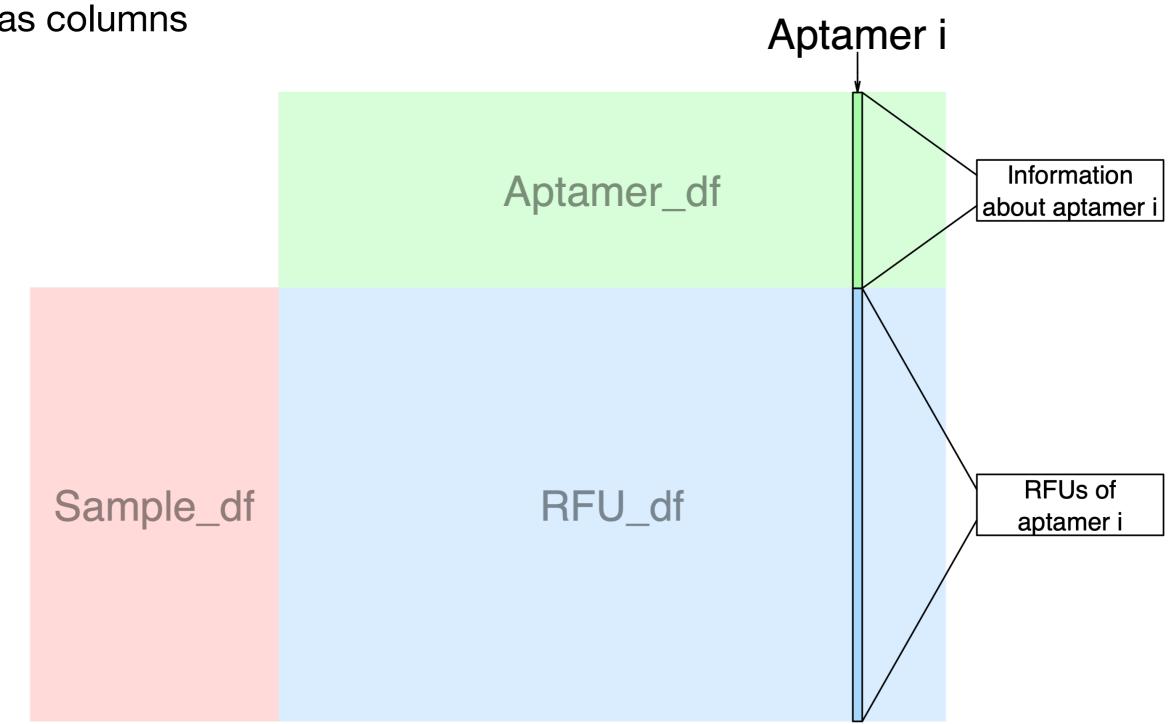
3 DataFrames in 1



Samples as Rows



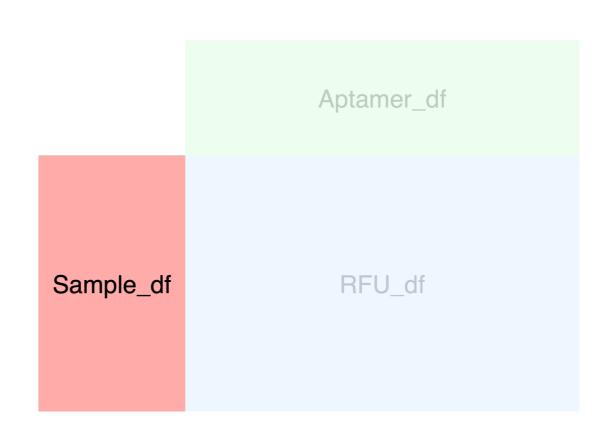
Aptamers as columns



Information on samples unrelated to aptamers

Sample_df

- SubjectID (VAP12345-78)
- Plateld, PlateRunDate, etc,....
- RowCheck (PASS/FLAG ?)
- +~30 others...



Information on aptamers unrelated to samples

Aptamer_df < MultiIndex >

• Aptamer ID (SomaId, unique)

• Protein IDs
Uniprot
EntrezGeneID

• Calibration values
• Dilution ratios
• Etc,...

WARNING#1:

1 aptamer → 1 protein

1 protein → X aptamer(s)

Different affinities to different protein conformations / states

Information on aptamers unrelated to samples

Aptamer_df < MultiIndex >

- Aptamer ID (SomaId, unique)
- Protein IDs

Uniprot

EntrezGeneID

- Calibration values
- Dilution ratios
- Etc,...

Aptamer_df

Sample_df

RFU df

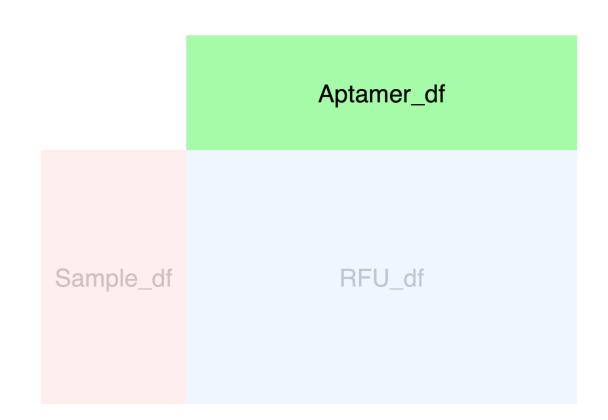
WARNING #2:

We **shouldn't** compare RFUs of **aptamer A and B** in patient X We **can** compare RFUs of aptamer A between **patient X and Y**

Information on aptamers unrelated to samples

Aptamer_df < MultiIndex >

- Aptamer ID (SomaId, unique)
- Protein IDs
 Uniprot
 - EntrezGeneID
- Calibration values
- Dilution ratios
- Etc,...



WARNING #3: Includes control aptamers!

You might want to filter:

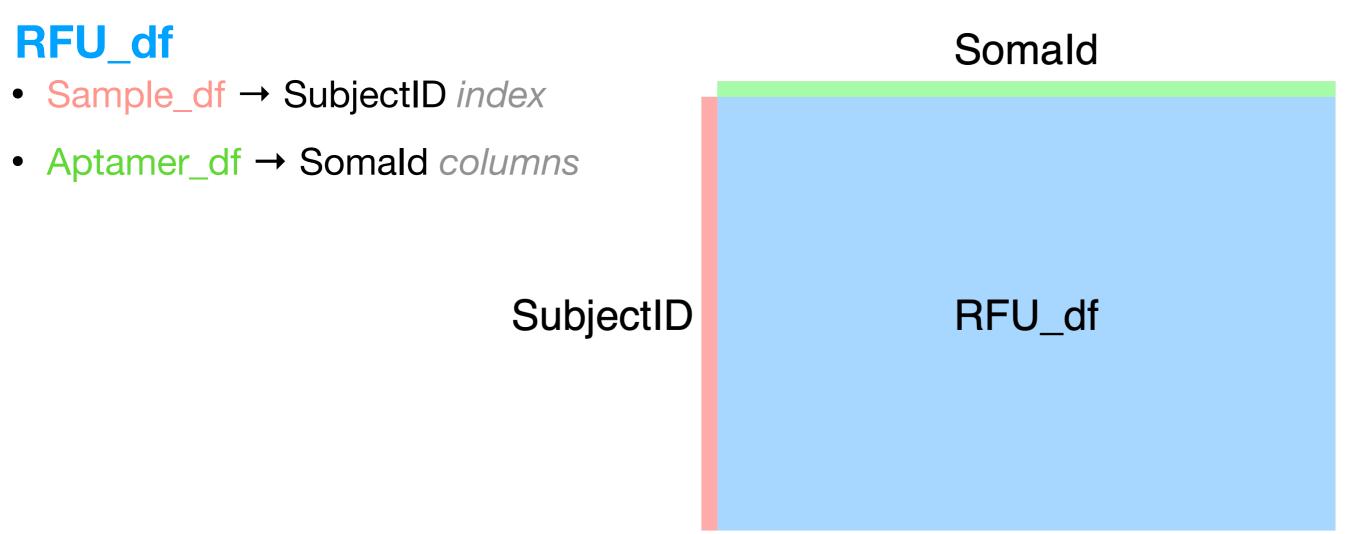
- 'Organism' == 'Human'
- 'Type' == 'Protein'

5284 → 4979 aptamers

Down to the essential...

Simplest solution IMO:

cut it back to a standard Pandas. Data Frame

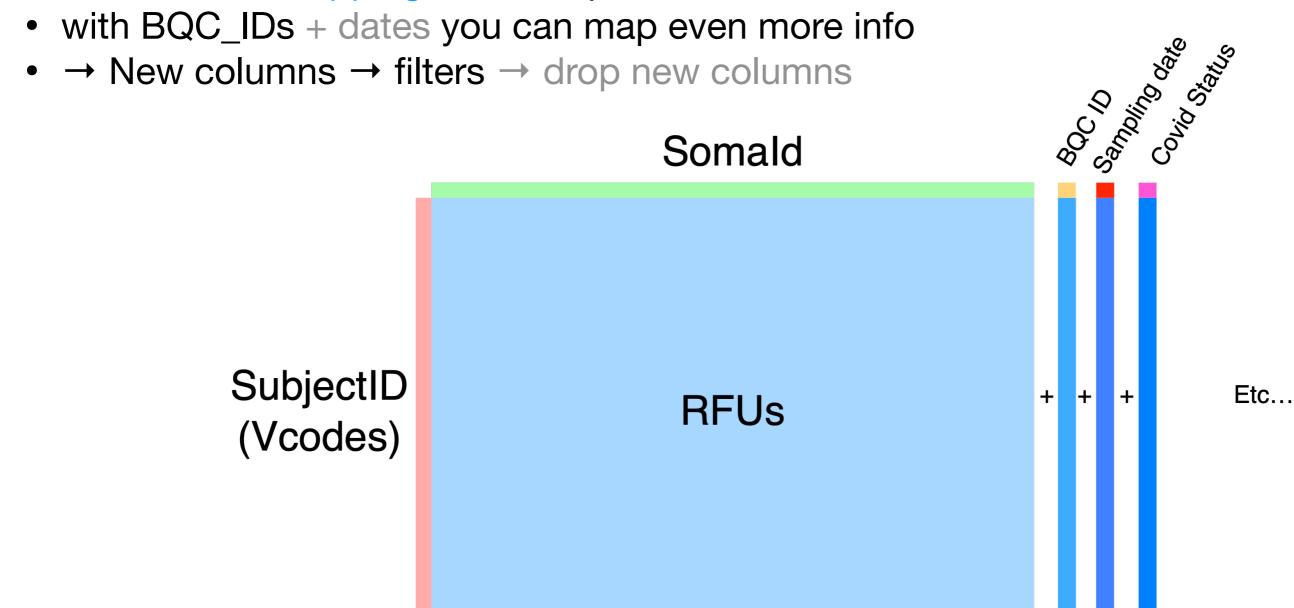


df_RFU=adat.pick_on_meta(axis=0, name='SampleType', values=['Sample']).pick_meta(axis=0, names=['SubjectID'])
df_RFU.columns=df_RFU.columns.get_level_values('Somald')

...and add whatever you need.

Then you can:

- Use aptamer_df to determine which Somalds are interesting
- Use the Vcode mapping file to map Vcodes to BQC_IDs, dates, etc,...



SomaScan - 1 dataset but 4 files

Couldn't be that simple, right?

There is actually 4 adat files available in BQC19 because:

- 1. Samples were processed in two separate batches
- 2. Each batch is available raw or already normalised by Somalogic

So 2 batches x 2 versions = 4 files

Our two cents:

Somalogic's normalisation method seemed sound so we used for the normalised versions

We log2 and z-score normalised (on columns) each batch separately before concatenating them to reduce batch effect.

Unsupervised Clustering

One way to use those data

Idea: comparing "mild" cases vs. "severe"
mixes together des heterogeneous profiles
which blurs any signal

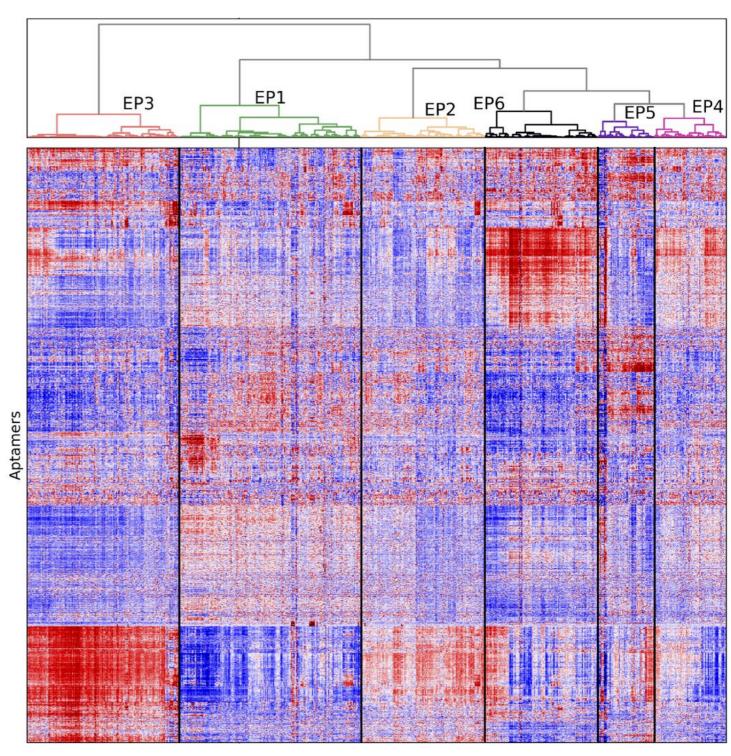
Ideally, we would like to separate those profiles and study their differences

1

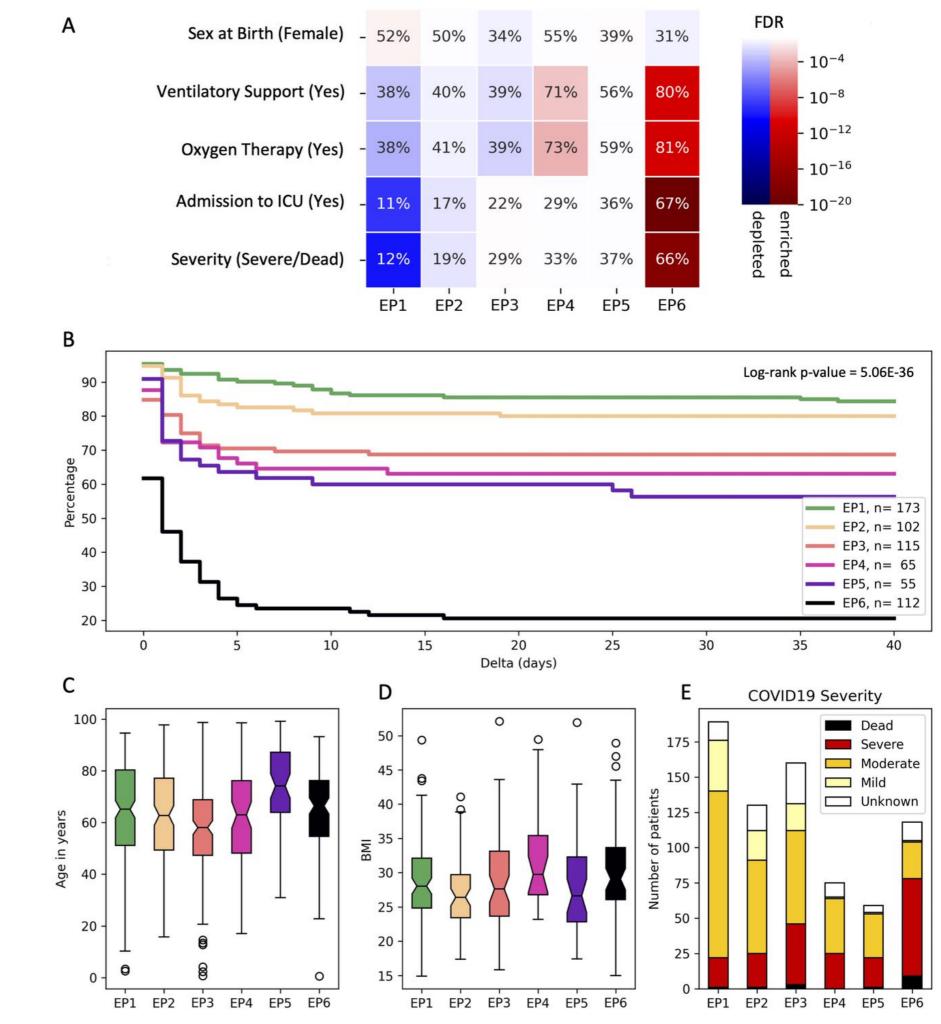
Unsupervised clustering of patients based on their protein expression only

(First blood draw)

731 patientsHospitalised **and** COVID positif



Unsupervised clustering of SARS-CoV-2 hospitalized patients identifies FGFR-signaling in severe COVID-19 acute respiratory distress syndrome

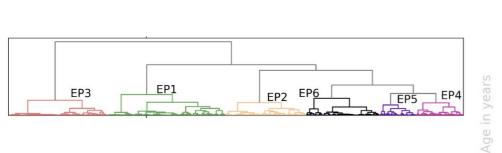


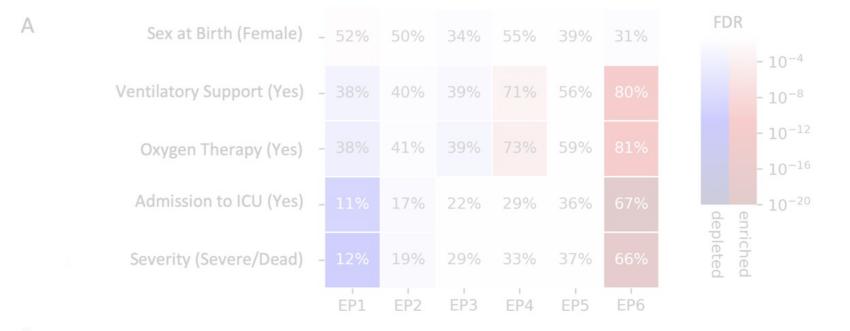
Kaplan-Meier

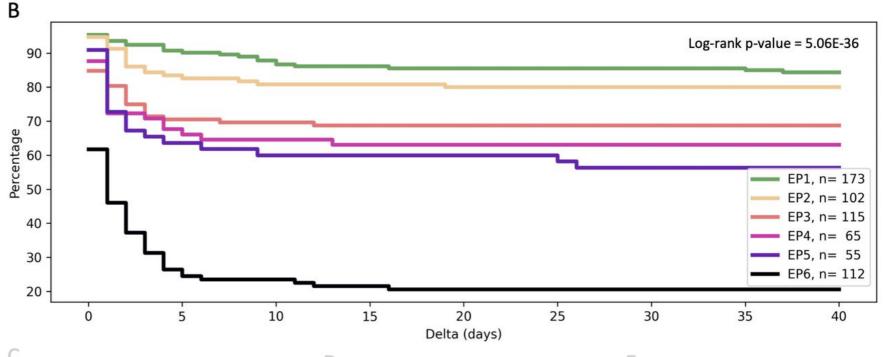
D0= admission to hospital E= admission to ICU (or death)

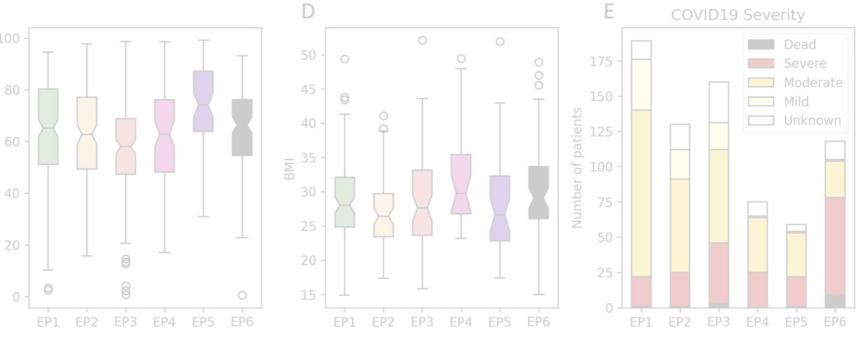
Clear deterioration of prognostic

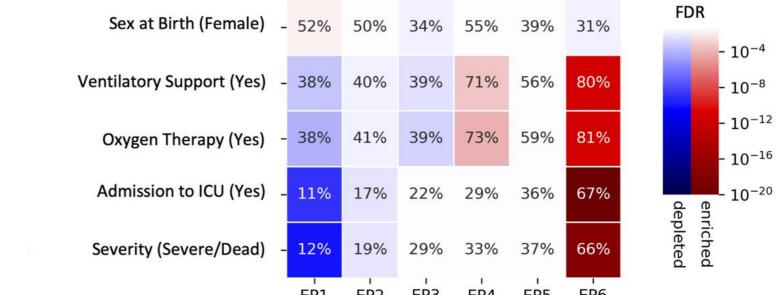
⇒ EPs' numbering





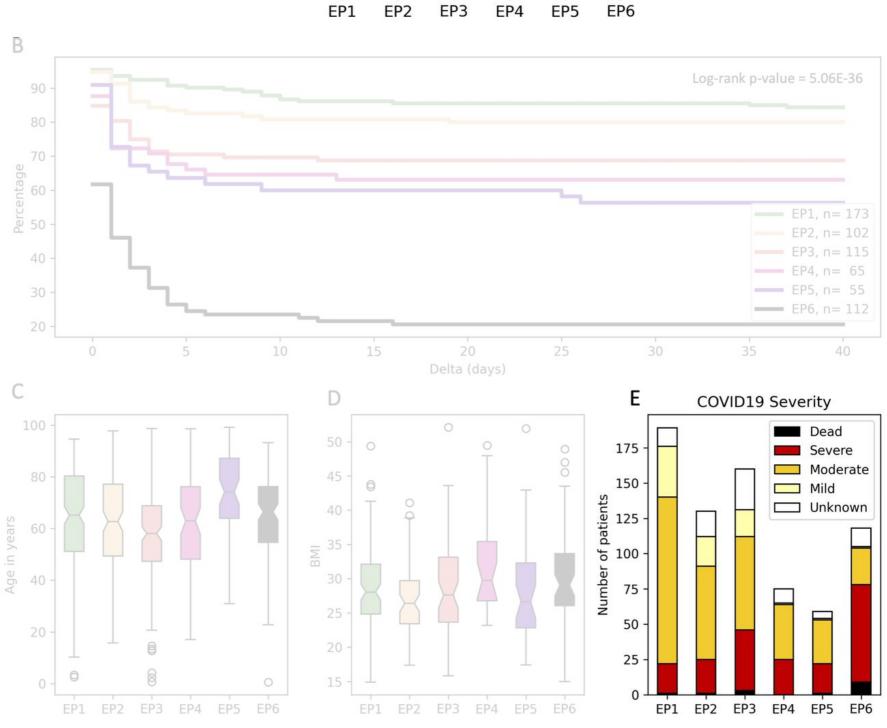






Clear deterioration of prognostic

EP1 and **EP6** as the two opposite extremes

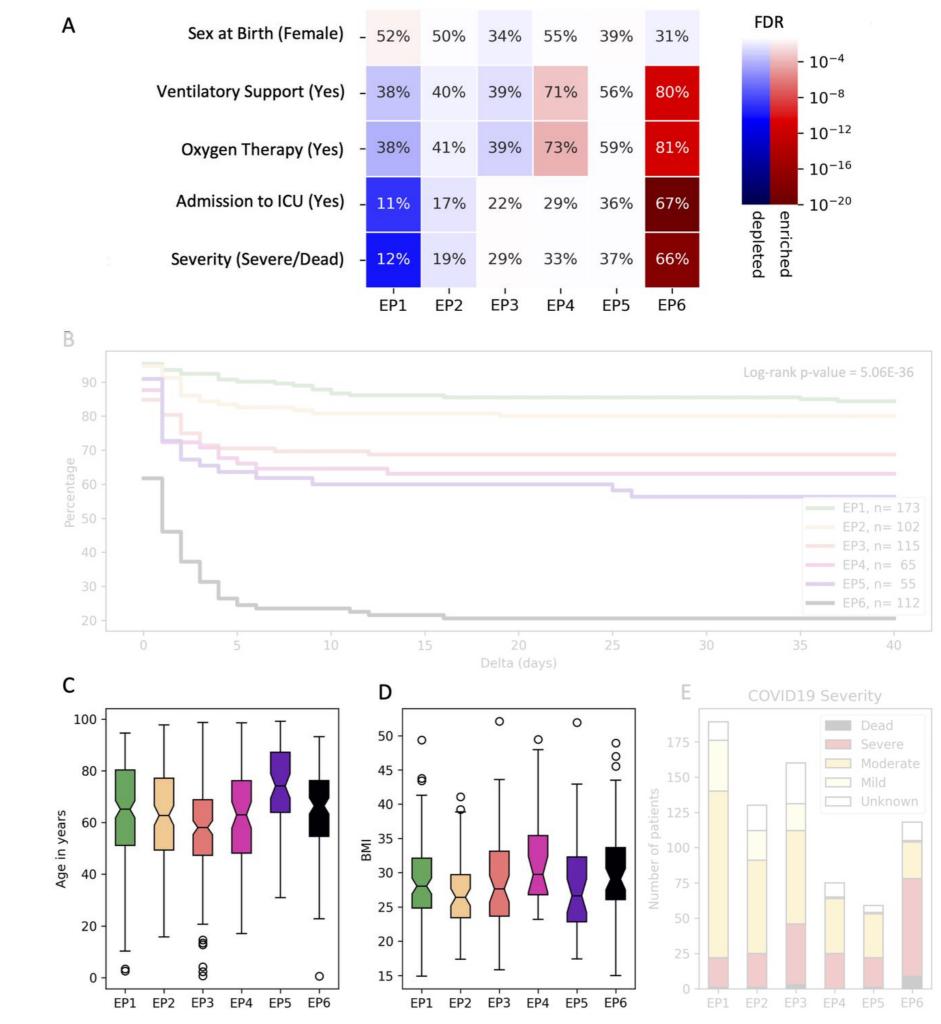


Weak connection with known suspects:

Sex: weak correlation (cf. EP3)

Age: also weak (cf. EP5)

BMI: unclear (missing data)



Thanks for your attention,

For the invitation,

And for the data!

Questions are obviously welcome!